SUPPLEMENTARY DATA

Figure S1: Fluorescent characterization of the reference surfaces. Fluorescence quantification of PDL and PLL-g-PEG determined by incubating FITC-PLL with non-fluorescently-labeled PLL-g-PEG and PLL-g-PEG-TRITC with PLL at the different ratios investigated. (n=3, mean ± standard error). Lines represent the expected fluorescence levels. The non-linearity of the data points can be attributed to the method of quantification where photobleaching of the fluorophores or background fluorescence from the surface are likely present. Despite the variations in the characterization, the trend can successfully be confirmed with this approach.
**Figure S2: Schematic of the assay format.** A 24-well plate illustrating microcontact printed stripe patterns of 18 coverslips of the experimental protein and 6 of the negative control, backfilled with various RS mixtures of low affinity PEG and high affinity PDL at different ratios across the plate.
Figure S3: Cell response to stripes of fibronectin patterned by incubation and by microcontact printing. (a) Alternating 125 µm wide stripes of patterned fibronectin either incubated (I, blue) or microcontact printed (P, black) on the substrate. C2C12 myoblasts were grown on the substrate backfilled with a 100:0 PEG: PDL RS and stained for filamentous actin (green). (b) Fibronectin deposition through incubation or through printing is shown to not be statistically significant (p=0.83>0.05) as per the percentage of cells on incubated and printed fibronectin (n=4, mean ± standard error). (c) High magnification image of a single cell grown on the substrates and stained for the focal adhesion marker paxillin (red) and filamentous actin (green). The dashed line indicates the separation between the stripes. Scale bars are (a) 50 µm and (c) 20 µm.